# Partitioning Variation in Measurements of Beef Carcass Traits Collected Using Ultrasound

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#### **ULTRASOUND**

- Method of measuring carcass traits
- Utilized since the 1950's
- Quick, relatively inexpensive, non-invasive
- Readily incorporated into multiple-trait genetic prediction



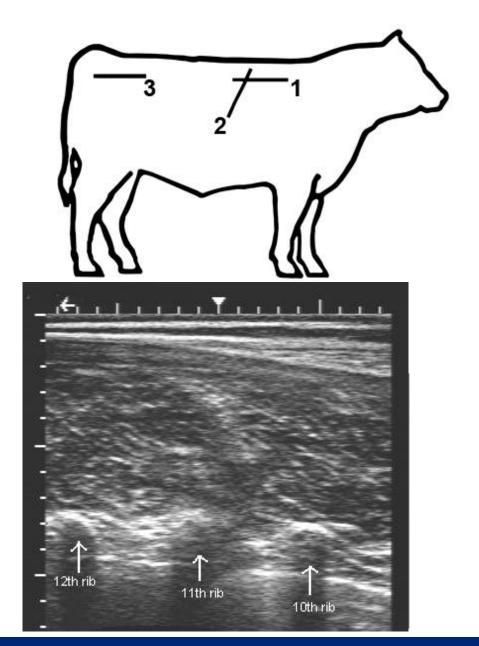
American Hereford Association

#### **CARCASS ULTRASOUND**

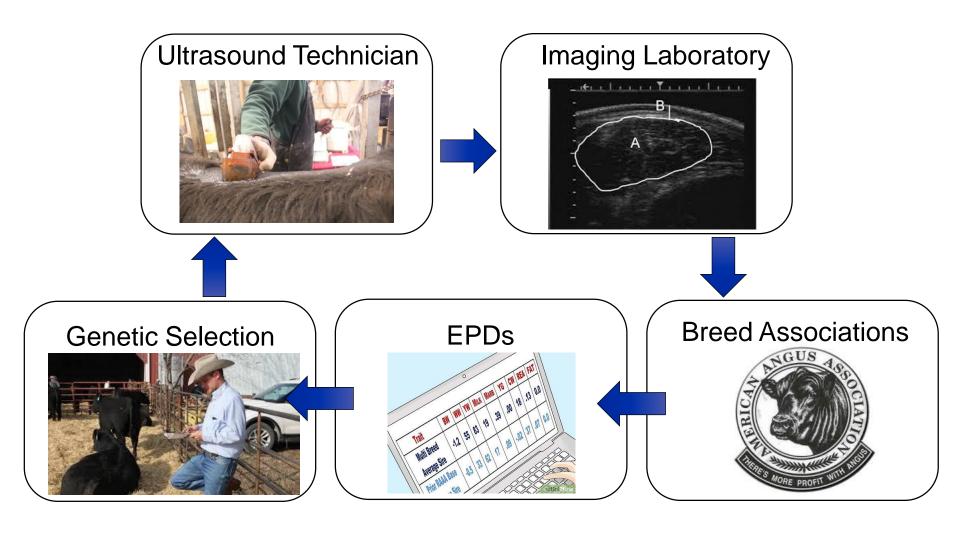
#### **Measurements**

- Intramuscular Fat (IMF)
- Longissimus Muscle Area
- Subcutaneous Fat
- Rump Fat

Top: University of Georgia Extension, 2018 Bottom: Carr et al., Ultrasound and Carcass Merit of Youth Market Cattle, University of Florida Extension



#### FLOW OF ULTRASOUND DATA



#### INTRODUCTION

- Abundant attention given to incorporation of data into systems of genetic evaluation
- Far less attention given to the underlying assumptions
- Technician and interpretive laboratory effects are assumed to be small due to UGC certification
- Homogeneity of additive genetic and residual variances

#### **HYPOTHESES**

- Homogeneity of within technician variances
- Technician variance = 0
- Homogeneity of additive genetic and residual variances across imaging laboratories
- Within trait, genetic correlations between imaging laboratories = 1

Informally, it does not matter who scans the cattle or which laboratory interprets the images



#### **DATA USED**







- Collected from 2015 to 2017
- Previously incorporated into national cattle evaluation
  - Animal ID
  - Contemporary group
  - Technician ID (includes technology)
  - Imaging laboratory

- Longissimus muscle area (LMA)
- Intramuscular fat (IMF)
- Subcutaneous fat depth (SFD)

All of the data came from images that had passed the QC of the interpretation laboratory and the breed association



#### **DESCRIPTION OF DATA - ANGUS**

	Interpretation	Number of scanning technicians - cor	N <sub>U</sub> ,	Phenotypic
Trait	Laboratory	$g_1$ N N=549	$N \approx 65$	5953 $3n$
LMA, cm <sup>2</sup>	1	61 2731	349	15.2
	2	14 - 1641	14719	16.3
	3	18 - 1415	16288	13.8
SFD, mm	1	61 - 2435	34952	2.77
	2	14 – 1641	14719	2.80
	3	18 – 1415	16288	2.72
IMF, %	1	61 - 2435	34960	1.30
	2	14 - 1641	14719	1.31
	3	18 – 1415	16288	1.51

#### **DESCRIPTION OF DATA - HEREFORD**

		Number of scanning technicians -	Phenotypic
Trait	Interpretation Laboratory	cor gro N N=4	572 \ N≈43158
LMA, cm <sup>2</sup>	1	45 - 221	23122 14.3
	2	12 - 1496	11490 15.5
	3	9 – 865	8546 13.9
SFD, mm	1	45 – 2214	21465 2.59
	2	13 – 1499	10366 2.51
	3	9 – 865	7914 2.71
IMF, %	1	45 – 2209	23120 0.98
	2	12 - 1498	11492 0.76
	3	9 – 867	8568 1.20

#### **DESCRIPTION OF DATA - SIMMENTAL**

Trait <sup>1</sup>	Interpretation Laboratory	Number of scanning technicians - conto	be	Phenotypic 48298
LMA, cm <sup>2</sup>	1	53_1703	25799	14.7
	2	11 - 780	6018	16.2
	3	23 - 1675	16481	15.6
SFD, mm	1	53 - 1963	25799	2.40
	2	11 - 780	6018	2.07
	3	23 - 1675	16481	2.34
IMF, %	1	53 – 1963	25799	1.02
	2	11 - 780	6018	0.81
	3	23 - 1675	16481	1.14

#### STATISTICAL MODEL

$$y_{ijk} = \mu + t_i + c_{ij} + a_{ijk} + e_{ijk}$$

 $y_{ijk}$  = Ultrasound carcass phenotype for  $k^{th}$  animal

 $\mu = Overall mean$ 

 $t_i = \text{Effect of } i^{th} \text{ technician}$ 

 $c_{ij} = \text{Effect of } j^{th} \text{ contemp. group scanned by } i^{th} \text{ technician}$ 

 $a_{ijk}$  = Effect of additive genetics by the  $k^{th}$  animal

 $e_{ijk}$  = Residual deviation from model effects

Linear model fitted using MTDFREML

All effects, except  $\mu$ , were considered <u>random</u>

$$t \sim N(0, \sigma_t^2)$$
  $c \sim N(0, \sigma_c^2)$   $a \sim N(0, A\sigma_a^2)$   $e \sim N(0, \sigma_e^2)$ 

#### MULTIVARIATE MODEL

$$\begin{bmatrix} y_1 \\ y_2 \\ y_3 \end{bmatrix} = \begin{bmatrix} 1\mu \\ 1\mu \\ 1\mu \end{bmatrix} + \begin{bmatrix} Z_1t_1 \\ Z_2t_2 \\ Z_3t_3 \end{bmatrix} + \begin{bmatrix} Z_4c_1 \\ Z_5c_2 \\ Z_6c_3 \end{bmatrix} + \begin{bmatrix} Z_7a_1 \\ Z_8a_2 \\ Z_9a_3 \end{bmatrix} + \begin{bmatrix} e_1 \\ e_2 \\ e_3 \end{bmatrix}$$

$$Var \begin{bmatrix} t_1 \\ t_2 \\ t_3 \end{bmatrix} = \begin{bmatrix} I\sigma_{t_1}^2 & 0 & 0 \\ 0 & I\sigma_{t_2}^2 & 0 \\ 0 & 0 & I\sigma_{t_3}^2 \end{bmatrix}$$

$$Var\begin{bmatrix} t_1 \\ t_2 \\ t_3 \end{bmatrix} = \begin{bmatrix} I\sigma_{t_1}^2 & 0 & 0 \\ 0 & I\sigma_{t_2}^2 & 0 \\ 0 & 0 & I\sigma_{t_3}^2 \end{bmatrix} \qquad Var\begin{bmatrix} c_1 \\ c_2 \\ c_3 \end{bmatrix} = \begin{bmatrix} I\sigma_{c_1}^2 & 0 & 0 \\ 0 & I\sigma_{c_2}^2 & 0 \\ 0 & 0 & I\sigma_{c_3}^2 \end{bmatrix}$$

$$Var\begin{bmatrix} e_1 \\ e_2 \\ e_3 \end{bmatrix} = \begin{bmatrix} I\sigma_{e_1}^2 & 0 & 0 \\ 0 & I\sigma_{e_2}^2 & 0 \\ 0 & 0 & I\sigma_{e_3}^2 \end{bmatrix}$$

$$Var \begin{bmatrix} a_1 \\ a_2 \\ a_3 \end{bmatrix} = \begin{bmatrix} A\sigma_{a_1}^2 & A\sigma_{a_1a_2} & A\sigma_{a_1a_3} \\ A\sigma_{a_2a_1} & A\sigma_{a_2}^2 & A\sigma_{a_2a_3} \\ A\sigma_{a_3a_1} & A\sigma_{a_3a_2} & A\sigma_{a_3}^2 \end{bmatrix}$$

SE of genetic correlations (Bijma and Bastiaansen, 2014)

## **RESULTS**



### Estimates of heritability assuming $\sigma_p^2 = \sigma_a^2 + \sigma_e^2$

Breed	Lab	LMA	SQF	IMF
Angus				
	1	$0.32 \pm 0.02$	$0.37 \pm 0.02$	$0.48 \pm 0.02$
	2	$0.27 \pm 0.03$	$0.33 \pm 0.03$	$0.67 \pm 0.04$
	3	$0.38 \pm 0.03$	$0.43 \pm 0.03$	$0.55 \pm 0.04$
Hereford				
	1	$0.35 \pm 0.02$	$0.26 \pm 0.02$	$0.34 \pm 0.02$
	2	$0.35 \pm 0.03$	$0.25 \pm 0.03$	$0.49 \pm 0.03$
	3	$0.34 \pm 0.03$	$0.29 \pm 0.03$	$0.42 \pm 0.03$
Simmental				
	1	$0.41 \pm 0.02$	$0.47 \pm 0.02$	$0.55 \pm 0.02$
	2	$0.45 \pm 0.05$	$0.41 \pm 0.05$	$0.52 \pm 0.05$
	3	$0.50 \pm 0.03$	$0.45 \pm 0.03$	$0.54 \pm 0.03$



#### Partitioning phenotypic variance of longissimus muscle area

	Variance components and percentages of phenotypic variance							ce
	$\sigma_a^2$	0/0	$\sigma_t^2$	0/0	$\sigma_{c:t}^2$	0/0	$\sigma_e^2$	%
Angus								
Lab 1	16.87	$7 \pm 1$	53.98	$23 \pm 4$	124.13	$54 \pm 3$	35.06	$15 \pm 1$
Lab 2	16.65	$6 \pm 1$	42.58	$16 \pm 6$	162.95	$61 \pm 4$	45.10	$17 \pm 1$
Lab 3	17.41	$9 \pm 1$	13.40	$7 \pm 3$	129.10	$68 \pm 2$	29.28	$15 \pm 1$
Hereford								
Lab 1	18.85	9 ± 1	34.24	$17 \pm 4$	120.75	$59 \pm 3$	30.50	$15 \pm 1$
Lab 2	20.45	$8 \pm 1$	15.57	$6 \pm 3$	169.03	$70 \pm 2$	35.97	$15 \pm 1$
Lab 3	14.75	$8 \pm 1$	8.14	$4\pm3$	143.16	$74 \pm 2$	28.11	$14 \pm 1$
Simmental								
Lab 1	27.31	$13 \pm 1$	57.21	$26 \pm 5$	93.89	$43 \pm 3$	38.60	$18 \pm 1$
Lab 2	33.35	$13 \pm 2$	60.64	$23 \pm 8$	126.81	$49 \pm 5$	40.31	$15 \pm 2$
Lab 3	30.57	$12 \pm 1$	49.98	$20 \pm 6$	133.84	$55 \pm 4$	30.67	$13 \pm 1$



#### Partitioning phenotypic variance of subcutaneous fat depth

	Variance components and percentages of phenotypic variance							nce
	$\sigma_a^2$	0/0	$\sigma_t^2$	0/0	$\sigma_{c:t}^2$	0/0	$\sigma_e^2$	0/0
Angus								
Lab 1	0.98	$13 \pm 1$	1.48	$19 \pm 3$	3.58	$47 \pm 2$	1.64	$21 \pm 1$
Lab 2	0.87	$11 \pm 1$	0.92	$12 \pm 5$	4.26	$54 \pm 3$	1.79	$23 \pm 2$
Lab 3	1.08	$15 \pm 2$	1.44	$19 \pm 6$	3.46	$47 \pm 4$	1.42	$19 \pm 2$
Hereford								
Lab 1	0.86	$13 \pm 1$	0.64	$10 \pm 2$	3.18	$47 \pm 2$	2.04	$30 \pm 1$
Lab 2	0.80	$13 \pm 2$	0.33	$5 \pm 3$	3.27	$52 \pm 2$	1.93	$31 \pm 2$
Lab 3	0.74	$10 \pm 2$	1.68	$23 \pm 9$	3.16	$43 \pm 5$	1.75	$24 \pm 3$
Simmental								
Lab 1	1.43	$25 \pm 2$	1.15	$20 \pm 4$	1.58	$28 \pm 2$	1.59	$28 \pm 2$
Lab 2	0.92	$22 \pm 3$	0.70	$16 \pm 6$	1.35	$31 \pm 3$	1.32	$31 \pm 3$
Lab 3	0.93	$17 \pm 2$	1.24	$23 \pm 6$	2.17	$39 \pm 3$	1.15	$21 \pm 2$



#### Partitioning phenotypic variance of percent intramuscular fat

	Va	Variance components and percentages of phenotypic variance						
	$\sigma_a^2$	0/0	$\sigma_t^2$	0/0	$\sigma_{c:t}^2$	0/0	$\sigma_e^2$	0/0
Angus								
Lab 1	0.34	$20 \pm 2$	0.43	$25 \pm 4$	0.56	$33 \pm 2$	0.37	$22 \pm 1$
Lab 2	0.52	$30 \pm 3$	0.21	$12 \pm 5$	0.73	$43 \pm 3$	0.26	$15 \pm 2$
Lab 3	0.51	$22 \pm 2$	0.33	$15 \pm 5$	1.03	$45 \pm 3$	0.41	$18 \pm 2$
Hereford								
Lab 1	0.16	$16 \pm 1$	0.21	$22 \pm 4$	0.37	$34 \pm 2$	0.27	$28 \pm 2$
Lab 2	0.15	$26 \pm 2$	0.07	$12 \pm 5$	0.23	$39 \pm 3$	0.13	$23 \pm 2$
Lab 3	0.24	$17 \pm 2$	0.20	$14 \pm 6$	0.69	$48 \pm 4$	0.32	$22 \pm 2$
Simmental								
Lab 1	0.28	$27 \pm 2$	0.27	$27 \pm 4$	0.26	$25 \pm 2$	0.23	$22 \pm 2$
Lab 2	0.17	$26 \pm 3$	0.10	$16 \pm 6$	0.22	$34 \pm 3$	0.16	$25 \pm 3$
Lab 3	0.31	$24 \pm 2$	0.18	$14 \pm 4$	0.55	$42 \pm 2$	0.26	$20 \pm 2$



#### LONGISSIMUS MUSCLE AREA

Estimates of genetic correlation and rank correlation of sires evaluated by pairs of interpretation laboratories (Number of sires)

		Lab 1	Lab 2	Lab 3
Angus				
	Lab 1		0.99 (417)	0.99 (501)
	Lab 2	$0.94 \pm 0.04$		0.99 (327)
	Lab 3	$0.96 \pm 0.04$	$0.94 \pm 0.04$	
Hereford				
	Lab 1		0.95 (245)	1.00 (199)
	Lab 2	$0.92 \pm 0.06$		0.96 (251)
	Lab 3	$0.98 \pm 0.06$	$0.88 \pm 0.06$	
Simmental				
	Lab 1		0.88 (341)	0.94 (510)
	Lab 2	$0.78 \pm 0.06*$		0.93 (320)
	Lab 3	$0.85 \pm 0.05$	$0.80 \pm 0.06*$	



#### SUBCUTANEOUS FAT DEPTH

# Estimates of genetic correlation and rank correlation of sires evaluated by pairs of interpretation laboratories (Number of sires)

		Lab 1	Lab 2	Lab 3
Angus				
	Lab 1		0.99 (418)	0.98 (501)
	Lab 2	$0.93 \pm 0.04$		0.98 (327)
	Lab 3	0.92 ± 0.04*	0.92 ± 0.04*	
Hereford				
	Lab 1		0.82 (232)	0.77 (185)
	Lab 2	$0.70 \pm 0.11$ *		0.49 (238)
	Lab 3	$0.58 \pm 0.14$ *	0.26 ± 0.14*	
Simmental				
	Lab 1		0.95 (341)	0.99 (510)
	Lab 2	$0.82 \pm 0.05 *$		0.93 (341)
	Lab 3	$0.94 \pm 0.04$	$0.79 \pm 0.06 *$	



#### PERCENT INTRAMUSCULAR FAT

Estimates of genetic correlation and rank correlation of sires evaluated by pairs of interpretation laboratories (Number of sires)

		Lab 1	Lab 2	Lab 3
Angus				
La	ab 1		0.99 (418)	0.99 (501)
La	ab 2	$0.95 \pm 0.03$		0.97 (327)
La	ab 3	$0.94 \pm 0.03*$	$0.89 \pm 0.03*$	
Hereford				
La	ab 1		0.97 (245)	0.97 (200)
La	ab 2	$0.89 \pm 0.06*$		0.93 (251)
La	ab 3	$0.87 \pm 0.07*$	$0.80 \pm 0.06 *$	
Simmental				
La	ab 1		0.94 (341)	0.97 (320)
La	ab 2	$0.79 \pm 0.05*$		0.96 (510)
La	ab 3	$0.88 \pm 0.04*$	$0.87 \pm 0.05*$	



#### **SUMMARY #1**

- Considerable variation among technicians; for all traits it is as large or larger than additive genetic merit
- Within technician estimates of variance are significantly heterogeneous (Bartlett's test) for all traits

#### **SUMMARY #2**

- Estimates of additive genetic variance are generally homogenous among the interpretation laboratories; but there may be exceptions
- Likewise, with exceptions the estimates of residual variance are generally homogenous among interpretation laboratories
- Genetic correlations among interpretation laboratories suggest that results reported from different laboratories may be slightly different "traits"; particularly for subcutaneous fat depth and IMF

#### RECOMMENDATIONS

- UGC should revisit the certification standards for both field technicians and image interpretation laboratories
- There may be merit in standardized methods of image interpretation that can be deployed across laboratories
- Breed associations should dive deeper into the data they receive, relative to carcass traits measured with ultrasound, to insure that they are meeting the BLUP assumptions of homogenous variance

#### **CLOSING THOUGHTS**

- There is work to do to make ultrasound the most valuable tool it can be for genetic improvement of beef cattle
- Data currently being collected using ultrasound technology is of unquestioned value in prediction of breeding values for carcass traits
- Rank correlations for sires having progeny with images interpreted in more than one laboratory indicate generally excellent agreement in their evaluations

#### **ACKNOWLEDGEMENTS**













**American Simmental Association** 

# Thank You



